### ADSORPTION OF HEAVY METALS FROM CONTAMINATED WATER ON Fe<sub>2</sub>O<sub>3</sub>/SiO<sub>2</sub> NANOCOMPOSITE AND NATURAL SILICA RICE HULL-BASED AND INVESTIGATE THEIR EFFECTS ON BLOOD BIOMARKERS AND RENAL TOXICITY IN RATS

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#### ABSTRACT

The detection of heavy metals in industrial effluent and some sectors of the global drinking water supply stands in direct opposition to the limits set by the regulatory authorities and the recommended Guidelines by the World Health Organization. Rice husk, as underutilized agricultural biomass, could be transferred into a promising low-cost adsorbent for removing heavy metals as it is abundant in nature, from a biogenic source, and requires little processing. In the present study, silica and Fe<sub>2</sub>O<sub>3</sub> silica nanocomposite were prepared from rice husk and used to clean contaminated water with a heavy metal mixture. Silica 0.5 and 1% exhibited up to 97.05 and 100% removal for Zn<sup>2+</sup>, Fe<sup>2+</sup>, and Ni<sup>2+</sup>, respectively, as they were affected by silica cristobalite amorphous microstructures and the non-uniform pore sizes. Silica 0.5% adsorbed more Mn<sup>2+</sup> (29.45 mg) and Fe<sup>2+</sup> (54.02 mg) than 1% silica. Fe<sub>2</sub>O<sub>3</sub>/silica nanocomposite 1% showed a specific selectivity to bind Hg<sup>2+</sup> with a maximum adsorption removal of 40.42% and adsorption capacity of 0.02 mg/ g. Serum urea, creatinine, and uric acid in rats exposed to drinking water cleaned with 0.5, 1% silica exhibited close  $p \le 0.05$  results to the normal values, and kidney histopathological examinations do not exhibit any sign of nephrotoxicity.

**KEY WORDS :** Rice hull ash, Silica nanocomposites, Water pollution, EDX-microelement composition, Adsorption capacity and efficiency, Kidney failure

#### **INTRODUCTION**

Heavy metals are elements having atomic weights from 63.5 to 200.6, and a specific gravity > 5.0, they are generally referring to metals and metalloids such as Manganese ( $Mn^{2+}$ ), Iron ( $Fe^{2+}$ ), Nickel ( $Ni^{2+}$ ), Mercury ( $Hg^{2+}$ ), and Lead ( $Pb^{2+}$ ) (Aliyah, 2012). The heavy metals are usually discharged from urbanization such as nuclear, metallurgical, tannery, mining, cosmetics, insecticides, photography, textiles, paints, dyes, and battery industries. They get accumulated both in underground and surface water, different parts of crops irrigated with contaminated water, and also in aquatic living animals (Ligate and Mdoe, 2015; Rehman *et al.*, 2018; Khan *et al.*, 2020). Small quantities of Fe<sup>2+</sup> and Mn<sup>2+</sup> (sometimes called trace elements) are essential for nutrition and health considerations. However, large concentrations of heavy metal did not degrade and accumulated in the living system, which ultimately leads to potential health risks and ecological disturbances. Moreover, prolonged exposure to heavy metals may substitute calcium by Pb<sup>2+</sup>, zinc by Cd<sup>2+</sup> and trace elements by Al<sup>2+</sup>. Furthermore, heavy metals react with oxygen and chloride compounds and expend their toxic effects in the human system (Aliyah, 2012; Rehman *et al.*, 2018). Pb<sup>2+</sup> can cause headache, irritability, cognitive and

\***Corresponding author's email:** soheirkenawy@yahoo.com, amalefsic@yahoo.com Amal M. H. Abdel-Haleem ORCID: <u>https://orcid.org/0000-0001-9912-3390</u> behavioral impairments, anemia, abdominal pains, blood pressure, nephritic damage, stomach and lungs cancers, gliomas, and reproductive system toxicity. Hg<sup>2+</sup> can cause many harmful health effects depending upon its form (metallic, inorganic, organic) and the level of exposure. At high levels of contamination, Hg2+ can cause nerve and kidney malfunctions and some neurotoxicity symptoms. Experimental animals exposed to mercury vapor showed obscure signs of pathological changes, cellular degeneration, brain necrosis, and suppressing natural killer cells. Meanwhile, Fe<sup>2+</sup> in drinking water after long-term exposure may cause nephritic disease, cancer, and anemia, along with metabolism disorders. Also, Mn<sup>2+</sup>in drinking water can cause Manganism and Alzheimer's. In addition, the Ni<sup>2+</sup> -sulfate and Ni<sup>2+</sup> chloride ingestion can cause severe cardiac arrest (Aliyah, 2012; Jaishankar et al., 2014; Abd El-Rahman et al., 2016; Reda et al., 2018; Nguyen, 2019).

Many experimental studies using animal trials showed a synergic effect between heavy metals exposure and renal failure associated with severe physiological and histological alterations. Also, the complete blood count (CBC) detected many abnormalities including, stress enzyme leakage into the bloodstream as affected by the redox reactions and the production of reactive oxygen species with a clear sign of oxidative damage in renal tissues (Abd El-Rahman *et al.*, 2016; Wasana *et al.*, 2017; Rehman *et al.*, 2018).

The detection of heavy metals in industrial effluent and some sectors of the global drinking water supply stands in direct opposition to the limits set by the regulatory authorities and the recommended Guidelines for Drinking Water Safety by the World Health Organization (WHO, 2006). That has raised concerns for the demand for water pollution control, especially with the intensification of global trends such as urbanization, deforestation, and climatic change (World Water Development Report, 2018). Therefore many efforts have been developed to remove heavy metals from water streams using various techniques, particularly the adsorption one. The adsorption technique is a promising technique of removal of heavy metals due to its high efficiency, easy operation, costeffectiveness, and environmental-friendly nature (Candido et al., 2021).

Rice husk is a by-product of the rice milling industry; it represents ~ 20% of whole milled rice. Internationally, the annual production of rice husk is ~ 100 million tons (Ong et al., 2019), In Egypt, rice production is estimated at 4.5 million tons (FAOSTAT, 2019), and then the volume of rice husk is about one million tons with a silica content of about 10%. However, this volume of rice husks is far in more than any local uses and poses environmental and disposal problems. It would be added value to transform this rice husk as underutilized agricultural biomass into a promising low-cost adsorbent, as it is abundant in nature, from a biogenic source, and requires little processing (Vieira et al., 2014; Ligate and Mdoe, 2015). The rice hull ash could be obtained after burning rice husk at 600° C with a silica (SiO<sub>2</sub>) content of 94.5–96.34% (Masoud et al., 2016). It is worth mentioning that the mineral ash SiO<sub>2</sub> is a promising adsorptive material due to its porous structure (10-75 µm) and large surface area (50,000 m<sup>2</sup>/kg) with high affinity and selectivity towards heavy metals; this is because of the presence of binding groups on its surface (Siddique and Cachim, 2018; Nguyen, 2019). It's worth more producing silica nanocomposite from rice ash SiO<sub>2</sub> to increase the crystallinity and the structural integrity and to be more economical and environmentally alternative than synthesized or commercial silica nanoparticles, which may not be sustainable and not match the cost requirement (Wang et al., 2012; Dang et al., 2017). Silica nanocomposite can be prepared by the sol-gel methodology from rice husk ash using metal oxides such as  $Fe_2O_3$  as supported oxide. The  $Fe_2O_3$  is a more attractive alternative adsorbent for water treatment as it is cost-effective and non-toxic. The physicochemical properties of the Fe<sub>2</sub>O<sub>2</sub>/SiO<sub>2</sub> nanocomposite are superior and can be used as adsorbents for a wide variety of reactions (Nguyen et al., 2019 8; Ramutshatsha-Makhwedzha et al., 2019; Agi et al., 2020).

Accordingly, the present research work were aimed to (i) transfer rice husk, as underutilized agricultural biomass, into promising adsorbents: silica and  $Fe_2O_3$  silica nanocomposite at concentrations of 0.5, 1%, for the cleaning of contaminated water with a heavy metal mixture ( $Mn^{2+}$ ,  $Fe^{2+}$ ,  $Ni^{2+}$ ,  $Hg^{2+}$ , and  $Pb^{2+}$ ) against Tetraethyl orthosilicate (TEOS) and  $Fe_2O_3$  / TEOS nanocomposite; (ii) assign the adsorption efficiencies and capacities of the prepared adsorbents via the scanning electron microscopy (SEM) and energydispersive X-ray spectroscopy (EDX) and then (iii) use an animal trial to investigate the health effects of different water treatments on blood biomarkers and renal histology of male rats.

#### MATERIALS AND METHODS

#### Materials

The rice hull was obtained from the Experimental Farm of Sakha Agricultural Research Station, Agricultural Research Centre, Kafr El Sheikh Governorate, Egypt. Synthetic or commercial silica: Tetraethyl orthosilicate (TEOS) with purity e"99.0% (GC) was purchased from Sigma-Aldrich, St. Louis, MO, USA. Iron (III) chloride hexahydrate FeCl3-6H<sub>2</sub>O, Lead (II) acetate trihydrate Pb (CH<sub>2</sub>COO)<sub>2</sub>. 3 H<sub>2</sub>O, Manganese (II) chloride tetrahydrate MnCl<sub>2</sub> ·  $4H_2O_1$ , Nickel(II) chloride hexahydrate NiCl<sub>2</sub> ·  $6H_2O_1$ and Mercury(II) chloride HgCl2 were obtained from Fluka Chemicals Corporation, St. Louis, USA. Distilled water was purified using the Millipore water purification system, Bedford, MA, USA. All other chemicals and reagents were of analytical grade.

## Preparation of natural silica and Fe<sub>2</sub>O<sub>3</sub> silica and Fe<sub>2</sub>O<sub>3</sub> TEOS nanocomposites

The rice hull biomass was cleaned and rinsed with purified water, dried at 155°C for 48 h, and then furnace for five hours at 600°C to get rice hull ash. Natural silica was prepared from rice hull ash using alkali solubilization, followed by acid precipitation according to the method described by Kamath and Proctor (1998). The rice hulls were dissolved in 1 M NaOH solution and boiled for one hour in a covered Erlenmeyer flask with constant stirring. The solution was flittered then a sol-gel method was performed to prepare silica nanocomposite using SiO<sub>2</sub>/ Fe<sub>2</sub>O<sub>3</sub> ratio of 70:30 at pH < 2 to prevent precipitation of Fe<sub>2</sub>O<sub>3</sub>. The obtained xerogels were dried at 100 °C and furnace at 600 °C for one hour. The TEOS nanocomposite was prepared by mixing the TEOS with an aqueous solution of FeCl<sub>3</sub>· 6H<sub>2</sub>O in acidified water (HCl) containing EtOH, according to Hrianca et al. (2000). The TEOS solution was filtrated then the above-mentioned sol-gel method described by

Kamath and Proctor (1998) was performed to obtain the TEOS nanocomposite.

#### Scanning electron microscopy (SEM)

The microstructure of natural, synthetic, and nanocomposite silica was examined by a scanning electron microscope JEOL, JSM-5200, Tokyo, Japan. The samples were sputter-coated with gold at a vacuum evaporator from 5 to 15 kV accelerating voltage and magnification power of 750-6,000×.

## Preparation of the drinking water contaminated with heavy metals

The drinking water contaminated with heavy metals, which included Mn2+, Fe2+, Ni2+, Hg2+, and Pb2+, was prepared by dividing the molecular weight of heavy metal compound by the molecular weight of each metal according to Nwokocha et al. (2010). To avoid precipitation, each heavy-metal was separately dissolved in 1000 mL of purified distilled water as a stock solution. Before use, 200 mL of this solution was mixed with 20 L purified distilled water to obtain 20 L of contaminated water contained the sum of five heavy metals at final working concentrations illustrated in Table 1. Five liters of the working solution were cleaned using each of 2.5; 5 g pre-activated (120° C for 2 hr) natural, synthetic, and nanocomposite silica. The drinking solutions were settled for 24 hr, and then filtrated and used for microelement analysis and biological assay. The solutions were freshly prepared each week to avoid heavy metals precipitation.

#### Energy-dispersive X-ray spec-troscopy (EDX)

Different water samples (purified, contaminated, and cleaned with natural, synthetic, and nanocomposite silica) were exposed to the microelement analysis using the Energy-dispersive X-ray spectroscopy (EDX) to detect the heavy quantitatively. The EDX model was INCA Sight Oxford equipped with a scanning electron microscope (JEOL, JSM-5500 LV). The mode of

Table 1. The contents and concentrations of the heavy metals in the contaminated drinking water

Compound	M. wt compounds (g/ mol)	M. wt heavy metal (g/ mol)	weight (g)	Final concentrations (mg/ L)
MnCl <sub>2</sub> · 4H <sub>2</sub> O	197.92	54.94	3.60	1000
FeCl3· 6H2O	270.30	55.85	4.84	1200
NiCl2 · 6H2O	237.69	58.69	4.05	100
HgCl,	271.5	201	1.35	10
Pb (CH <sub>3</sub> COO) <sub>2</sub> . 3 H <sub>2</sub> O	379.33	207.2	1.83	100

analysis was Window Integral, the detection limit was 0.1%, scan time was 200 seconds, and scan dimensions were  $20 \times 300 \times 250 \mu m$ .

# The adsorption capacity and efficiency of natural, synthetic, and nanocomposite silica for heavy metals

The adsorption efficiency (AE) for heavy metals with natural, synthetic, and nanocomposite silica was calculated in percentages (%) at fixed conditions at  $37\pm2^{\circ}$  C after 24 hr of water cleaning following equation 1:

$$AE\% = \frac{(C-T)}{C} \times 100$$
 ... (1)

Where C is the concentrations of heavy metals (mg/l) in contaminated water; T is the concentrations of the heavy metals (mg/l) after the water cleaning with natural, synthetic, and nanocomposite silica.

The adsorption capacity (AQ), which refers to the amount of heavy metal absorbed per unit mass of natural, synthetic, and nanocomposite silica, was calculated (mg/g) at fixed conditions at  $37\pm2^{\circ}$  C after 24 hr of water cleaning following equation 2.

$$AQ(mg/g) = \frac{(C-T)V}{m} \times 100 \qquad ...(2)$$

Where C is the concentrations of heavy metals (mg/l) in contaminated water; T is the concentrations of the heavy metals (mg/l) after 24 of the water cleaning; V is the volume of the solution (L); m is the mass of adsorbent 5 and 10 g).

#### **Animals and Experimental Design Protocol**

Eight-week- male Wistar rats weighing 180±20 g were obtained from Experimental Laboratory Animal Unit, Food Technology Research Institute, Agriculture Research Center, Al Giza, Egypt. The animals were kept at constant room temperature 25±5° C with 12 h of light/ dark cycles. During the acclimatization period, all rats had access to distilled water *ad libitum* and were received a standard laboratory chow prepared with the recommended daily dose for each nutrient according to the National Research Council (1995). Then rats were divided into ten homogenous groups, each of six, and then exposed to oral administration of different water treatments for ten weeks according to the following protocol:

- Group 1: fed normal rat chow+ purified distilled water *ad libitum*
- Group 2: fed normal rat chow + contaminated

drinking water with heavy metals.

- Group 3: fed normal rat chow + 0.5% natural silica SiO<sub>2</sub> + contaminated drinking water with heavy metals.
- Group 4: fed normal rat chow + 1% natural silica SiO<sub>2</sub> + contaminated drinking water with heavy metals
- Group 5: fed normal rat chow + 0.5% synthetic silica TEOS + contaminated drinking water with heavy metals.
- Group 6: fed normal rat chow+ 1% synthetic silica TEOS + contaminated drinking water with heavy metals
- Group 7: fed normal rat chow+ 0.5% natural  $Fe_2O_3/$  SiO<sub>2</sub> nanocomposite+ contaminated drinking water with heavy metals
- Group 8: fed normal rat chow+ 1% natural Fe<sub>2</sub>O<sub>3</sub>/ SiO<sub>2</sub> nanocomposite + contaminated drinking water with heavy metals
- Group 9: fed normal rat chow+ 0.5% synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite + contaminated drinking water with heavy metals
- Group 10: fed normal rat chow+ 1% synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite + contaminated drinking water with heavy metals

#### **Biochemical assays**

Rats were sacrificed, and blood samples were collected via cardiac puncture and separated into the serum. Serum urea, creatinine, and uric acid were quantified according to the manufacturer's instructions (Bio Diagnostic Company, Dokki, Al-Giza, Egypt).

#### Histopathological examination

The renal tissues were fixed in 10% neutral formaldehyde and dehydrated with a gradient sequence of aqueous EtOH from 70 to 100%. After that, the dehydrated tissues were embedded into the paraffin, cut into 4-5  $\mu$ m sections, and then stained with Hematoxylin dye (H&E Staining), according to Suvarna *et al.* (2018). Careful examination of the stained sections was made using a DM750 light microscope, Leica Microsystems IR GmbH, Switzerland, to identify the histopathological alterations induced by different treatments.

#### Statistical analysis

The data of this study were analyzed using

computer software Jandel Sigma Stat Statistical Software version 2.0 for Windows. An analysis of variance (ANOVA) using a One- Way completely randomized design followed by Fischer's test at  $P \le 0.05$ , to compare between means.

#### **RESULTS AND DISCUSSION**

## The Morphological structure of natural, synthetic, and nanocomposite silica

Figure 1 (a-d) represents the morphological structures of natural, synthetic, and nanocomposite silica. The natural silica (Fig. 1a) exhibited a cristobalite amorphous form with traces of crystalline quartz. The non-uniform particle size was due to the high burning temperature of the rice hull (Thiedeitz *et al.*, 2020). In addition, some particles can be seen agglomerated and stuck to each other due to the high surface energyand free silanol groups (Si–OH) on the natural silica surface (Basri *et al.*, 2020). The high concentrations of Si–OH groups coupled with ordered cristobalite amorphous structures, and the non-uniform pore sizes, make natural silica efficient sorbent for heavy

metals in polluted water (Hu et al., 2020). After solgel preparation, the natural Fe<sub>2</sub>O<sub>2</sub>/ SiO<sub>2</sub> nanocomposite (Fig. 1b) was not relatively different from that of the original natural silica. The microstructure became immense irregular, with expander particle size distribution and more agglomerations (Palanivelu et al., 2016 30). The agglomeration referring to the aggregation of Fe<sub>2</sub>O<sub>3</sub> on its surface as a result of Van der Waals forces and later that may decrease the adsorption efficiency of Fe<sub>2</sub>O<sub>2</sub>/ natural SiO<sub>2</sub> nanocomposite the (Ramutshatsha-Makhwedzha et al., 2019). The synthetic TEOS (Fig. 1c) exhibited silica sand flaky aggregates with sub-angular to angular shapes and sharpened edges. Moreover, the microstructure of the synthetic TEOS showed less mount of agglomerations. After sol-gel preparation, the synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite (Fig. 1d) exhibited a fully modified and uniform crystalline structure. This structure is beneficial for the surface activity of the composite where it increases the surface to volume ratio, eventually increase the interaction with heavy metal molecules. In our previous work (Reda et al., 2018), we analyzed the



**Fig. 1.** Scanning electron microscope (SEM) images of natural, synthetic, and nanocomposite silica (**a**): natural silica; (**b**): natural Fe<sub>2</sub>O<sub>3</sub>/ SiO<sub>2</sub> nanocomposite; (**c**): synthetic TEOS (**d**): synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite.

porosity by nitrogen adsorption/desorption isotherm using the BJH. We found that the surface area of natural silica nanocomposite was  $(43.12 \text{ m}^2/\text{ g})$  smaller than the synthetic one  $(320 \text{ m}^2/\text{ g})$ . So we can expect a decrease in the interaction between natural silica nanocomposite and heavy metals.

The EDX elemental composition of the drinking water and the adsorption efficiency (AE) and the adsorption capacity (AQ), of the natural, synthetic, and nanocomposite silica for heavy metals

Table 2 displays the EDX elemental composition (mg/l) in the prepared drinking water before and after cleaning with 0.5, 1% natural, synthetic, and nanocomposite silica. The general trend of the EDX elemental composition (mg/l) is as shown contaminated water > natural Fe<sub>2</sub>O<sub>3</sub>/ SiO<sub>2</sub> nanocomposite (0.5%) > synthetic silica TEOS (0.5%) > natural Fe<sub>2</sub>O<sub>3</sub>/ SiO<sub>2</sub> nanocomposite (1%) > synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite (1%) > synthetic silica TEOS (1%) > natural silica SiO<sub>2</sub> (0.5%) > natural silica SiO<sub>2</sub> (1%) > purified water. It can be inferred that the lowest  $P \le 0.05$  composition of Mn<sup>2+</sup>, Fe<sup>2+</sup>, Ni<sup>2+</sup>, Hg<sup>2+</sup>, and Pb<sup>2+</sup> were found in contaminated water treated with natural silica SiO<sub>2</sub>

(1%, 0.5%), however the highest  $P \le 0.05$  elemental composition were found to be in the  $Fe_2O_2/SiO_2$ nanocomposite (0.5, 1%), followed by synthetic TEOS (0.5%). These results could be explained more clearly within the amounts of heavy metal ions removed, after water cleaning, per unit mass of natural, synthetic, and nanocomposite silica (Figure 2 and 3). Adsorbent concentration is an important parameter in the adsorption of heavy metal ions from contaminated water owing to its effects on the amounts of heavy metal ions removed per unit mass of adsorbent (Kukwa et al., 2020). In the present work, the percentages of heavy metals removed were increased with the increasing mass of the natural, synthetic, and nanocomposite silica from 5 to 10 g (Fig. 2). The natural silica SiO<sub>2</sub> at 0.5 and 1% exhibited efficient removal up to 97.05%, and complete removal of 100% for Zn<sup>2+</sup>, Fe<sup>2+</sup>, and Ni<sup>2+</sup>, respectively, as it affected by its superior microstructure (Fig. 1 a) with high concentrations of Si–OH groups coupled with ordered cristobalite amorphous structures, and the non-uniform pore sizes, make natural silica efficient sorbent for complete removal of Zn<sup>2+</sup>, Fe<sup>2+</sup>, and Ni<sup>2+</sup> (Basri et al., 2020). Also, synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite



Fig. 2. Adsorption efficiency (AE, %) for heavy metals of the natural, synthetic, and nanocomposite silica

<b>Fable 2.</b> EDX	elemental	composition (m	g/ L) in the pre	pared drinking	water before an	d after cleaning	; with natural, sy	mthetic, and na	nocomposite si	lica
Heavy	Purified	Contaminated	Nature	al silica	Synthet	tic silica	Natural F	e2O3/ SiO2	Synthetic F	e2O3/ TEOS
metals	water	water	ŝ		TE	SOS	nanoco	mposite	nanoco	mposite
			0.5%	1%	0.5%	1%	0.5%	1%	0.5	1%
Mn	ND	151.73±0.3ª	4.47±0.95	ND	$117.94\pm1.1^{b}$	58.5±2.1 <sup>d</sup>	$146.97\pm0.64^{a}$	$141.6\pm0.51^{ab}$	$107.97\pm 2.0^{\circ}$	100.6±1.25°
Fe	ND	280.20±5.2ª	10.08±0.23°	ND	$239.48\pm 1.6^{ab}$	$142.36\pm 1.8^{d}$	$206.64\pm1.8^{b}$	$204.72\pm5.3^{b}$	$172.08\pm1.4^{\rm cb}$	$142.24\pm0.15^{d}$
Ni	ND	$2.35\pm1.4^{a}$	$0.7\pm0.6^{d}$	ND	$1.59\pm0.49^{\mathrm{ab}}$	1.03±0.63°	$1.84\pm0.18^{\mathrm{ab}}$	$1.06\pm0.84^{\circ}$	$1.73\pm0.69^{ab}$	$1.60\pm0.6^{\mathrm{ab}}$
Hg	ND	$0.48\pm0.64^{a}$	$0.46\pm1.5^{a}$	$0.41\pm0.52^{a}$	$0.46{\pm}1.6^{a}$	$0.45\pm0.13^{a}$	$0.41\pm0.44^{a}$	$0.28\pm0.16^{b}$	$0.47\pm0.88^{a}$	$0.46\pm0.53^{a}$
Pb	ND	$10.22\pm3.0^{a}$	$8.65\pm 1.1^{b}$	$7.24\pm1.37^{d}$	$9.65{\pm}0.94^{\rm ab}$	$9.19{\pm}0.94^{ m b}$	$9.93\pm1.1^{ab}$	$9.81\pm0.99^{ab}$	$9.28\pm2.3^{b}$	8.07±0.8°
ND: not detec	ted.									
Data are prese	ented as m	eans ± SDM (n=ć	3) & Means wit	thin a column w	vith different lett	ers are significa	intly different at	$P \le 0.05$ .		
Table 3. Seru	m biomark	ers of rat kidney	y treated with t	he prepared dri	inking water.					
Serum	Purifie	d Contam	uinated	Natural silic	ä	Synthetic silica	Natu	ral Fe2O3/ SiO2	Synthetic	Fe2O3/ TEOS
biomarkers	wateı	wa	ter	$SiO_2$		TEOS	nai	nocomposite	nanoc	composite
			0	).5% 1	% 0.5%	% 1%	0.5%	1%	0.5	1%

 $\leq 0.05$ . പ different at SDM (n=5) & Means within a column with different letters are significantly Data are presented as means ± at 0.5 and 1% was more efficient than natural Fe<sub>2</sub>O<sub>3</sub>/ SiO<sub>2</sub> nanocomposite in removing  $Mn^{2+}$ ,  $Fe^{2+}$ ,  $Ni^{2+}$ , and  $Pb^{2+}$ . The maximum percentages were 33.70, 49.24, 31.79, and 21.01%, respectively. These results owing to that the surface area of Fe<sub>2</sub>O<sub>3</sub>/ SiO<sub>2</sub> nanocomposite (Fig.1b) was decreased as affected by the aggregation of Fe<sub>2</sub>O<sub>3</sub> and the Van der Waals forces (Ramutshatsha-Makhwedzha et al., 2019). In our early study (Reda et al., 2018), we found that the surface area of natural silica nanocomposite was  $(43.12 \text{ m}^2/\text{ g})$  smaller than the synthetic one  $(320 \text{ m}^2/\text{ g})$ . But what worth that the natural Si<sub>2</sub>O<sub>2</sub> is nanocomposite 1% has specific priority and selectivity to bind certain types of metal depending upon above- mentioned aggregations and Van der Waals forces (Aliyah, 2012), where it showed the highest efficiency for removing Hg<sup>2+</sup> with a maximum percentage of 40.42%.

The results in Figure 3 exhibited a reverse relationship-the adsorption capacity (AQ, mg/g) was decreased when the silica mass increased from 5 to 10 g. For example; 0.5% natural silica adsorbed more Mn<sup>2+</sup> (29.45 mg/g), and Fe<sup>2+</sup> (54.02 mg/g) than  $Mn^{2+}$ (15.17 mg/g), and Fe<sup>2+</sup>(28.02 mg/g) for 1% natural silica. This result was due to the saturation of accessible active sites on sorbent materials above a certain concentration of heavy metals (Gorzin et al., 2018; Abdel Halim et al., 2019). It can be inferred that the higher composition of coexisting heavy metals in the contaminated water resulted in the stronger competition among heavy metals, where the elemental composition of Fe<sup>2+</sup> (280.20±5.2 mg/L) was higher than Ni<sup>2+</sup> (2.35 $\pm$ 1.4), and Hg<sup>2+</sup>  $(0.48\pm0.64)$  (Table 2), so Fe<sup>2+</sup> could compete with Ni<sup>2+</sup> and Hg<sup>2+</sup> to be adsorbed more on the surface of the natural silica. Another reason may be related to the ionic radius of heavy metals, where the atomic radius of Fe (0.126 nm) < Hg (0.3150 nm) < Ni (0.3163 nm),the lower atomic radius, lower hydration energy of the heavy metals was easier for the adsorption (Ouyang et al., 2019).

#### Serum biomarkers for rat kidney function

Table 3 represents the serum biomarkers of

49.3±1.15<sup>d</sup> 1.36±0.1<sup>c</sup> 5.73±0.2<sup>c</sup>

51.6±1.15<sup>d</sup> 1.51±0.1<sup>c</sup>

 $55.6\pm1.15^{\circ}$  $1.76\pm0.1^{\circ}$  $6.8\pm0.1^{\circ}$ 

67.3±2.08<sup>b</sup>

41.6±1.15<sup>ef</sup> 1.06±0.1<sup>e</sup> 5.23±0.2<sup>d</sup>

45.3±1.15<sup>e</sup> 1.23±0.1<sup>cd</sup> 6.1±0.12<sup>c</sup>

39.0±1.0<sup>f</sup> 0.73±0.1<sup>f</sup> 3.66±0.2<sup>e</sup>

> 0.76±0.1<sup>f</sup> 3.76±0.2<sup>e</sup>

87.0±2.0<sup>a</sup> 2.2±0.2<sup>a</sup> 8.3±0.2<sup>a</sup>

39.3±0.57<sup>i</sup>

39.3±1.52<sup>6</sup> 0.76±0.1<sup>6</sup> 3.56±0.1<sup>e</sup>

> Creatinine Uric acid

Urea

 $1.86\pm0.1^{b}$ 

7.1±0.2<sup>b</sup>

6.2±0.3°

1595



**Fig. 3.** Adsorption capacity (AQ, mg/ g) for heavy metals of the natural, synthetic, and nanocomposite silica

rat kidneys fed the prepared drinking water. There were significant ( $p \le 0.05$ ) alterations in the levels of serum urea, creatinine, and uric acid in rats exposed to drinking water contaminated with heavy metals, compared to the rats exposed to purified water. On the other hand, both natural silica  $SiO_2(0.5, 1\%)$  and synthetic TEOS (1%) exhibited close results to the normal values observed in rats exposed to purified water. In comparison, synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite at concentrations 0.5 and 1% showed better results than Fe<sub>2</sub>O<sub>2</sub>/SiO<sub>2</sub> nanocomposites at the same concentrations. These results are coherent with the results in Table 2 and Fig. (1b & d), where synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite had a larger surface area (Fig. 1d) and higher efficiency in the adsorption of heavy metals than natural  $Fe_2O_3/SiO_2$ nanocomposite (Fig. 2), as well as natural silica SiO<sub>2</sub> had higher efficiency in the adsorption of heavy metals than Synthetic silica TEOS (Fig. 2).

There are controversial results regarding serum biomarkers in the experimental trials. A group of researchers (Oyagbemi *et al.*, 2015) found that oral administration of Pb (CH<sub>3</sub>COO)<sub>2</sub> in rats caused p < 0.01 increase in the blood urea and serum creatinine, most likely happened in acute and chronic intrinsic renal disease, and also when there was a decrease in the effective circulating blood volume with a

decrease in renal perfusion (Basile et al., 2012). While another group (Waghmare et al., 2015) found that serum urea was reduced in all rat groups treated with Ni<sup>+2</sup>, Fe<sup>+2</sup>, and Mn<sup>+2</sup>. Also, Andjelkovic et al. (2019) found that acute exposure to Cd<sup>+2</sup> and Pb<sup>+2</sup> administered alone or in a mixture form resulted in (p < 0.01) decreased urea concentrations, while there was a slight (p < 0.01) increase in creatinine levels compared to the control group. On the other hand, Wang et al. (2020) found no significant changes in the biochemical parameters included urea, creatinine, and uric acid in experimental animals fed a multi-heavy metal mixture with 500 mg/ kg body weight for six months, compared to untreated animals. The authors stated that the kidney might still be in a compensatory stage, while the heavy metals were accumulated in different degrees in the kidney.

#### Histology of rat kidney

Figure 4 (a-j) displays the histopathological sections of kidneys for male rats of various studied groups. The kidney section of the rats exposed to purified water (Fig. 4a) showed the normal structure of renal parenchyma, normal podocytes, normal glomerular capillaries with thin and delicate loops, normal number of the endothelial cells, and normal surrounding tubules. However, the kidney section of the rats exposed to contaminated water with heavy metals (Fig. 4 b) showed sever histopathological changes, including necrosis and degeneration of the renal tubules, interstitial fibrosis, hypertrophy, and hemorrhage (Albu et al., 2021). Hypertrophy is characterized by an increase in the Bowman's Space, expansion of the connective tissues, and shrank of the glomerulus. The hemorrhage is accompanied by vacuolization of the epithelial cells, a deposit of the RBCs and clogged blood vessels in the capillaries, and decreased hematopoietic tissue (Riaz et al., 2020). These histological changes could be explained as; heavy metals-induced nephrotoxicity is mediated by rapid selective accumulation of heavy metals and metalloid complexes which are filtrated by kidney glomerulus, taken up by the cytosolic fraction of



proximal tubular epithelium and leads to gradual loss of the kidney function and tissue damage (Abd El-Rahman *et al.*, 2016; Seif *et al.*, 2019).

The kidney section of the rats exposed to contaminated water treated with 0.5, 1% natural silica (Fig.4 c & d), and 1% synthetic TEOS (Fig. 4 f) do not exhibit any sign of nephrotoxicity either hemorrhage or tissue damage; they were most likely similar to that section for the rats exposed to purified water. Meanwhile, the kidney section of the rats exposed to contaminated water treated with 0.5% synthetic TEOS (Fig. 4 e) exhibited renal tissue completely normal, except for mild degeneration in the dilated capillaries and the lining epithelium of the proximal and distal tubules (Seif *et al.*, 2019).

The kidney section of the rats exposed to contaminated water treated with  $0.5\% \text{ Fe}_2\text{O}_3/\text{ SiO}_2$  nanocomposite showed cystic dilatation of renal





Fig. 4. Histological sections of renal rat exposed to different drinking water (Stained with H&E, 200×). (a) Purified water.
(b) Contaminated water with heavy metals. (c) Contaminated water treated with 0.5% natural silica. (d) Contaminated water treated with 1% natural silica. (e) Contaminated water treated with 0.5% synthetic TEOS. (f) Contaminated water treated with 1% synthetic TEOS. (g) Contaminated water treated with 0.5% Fe<sub>2</sub>O<sub>3</sub>/SiO<sub>2</sub> nanocomposite. (h) Contaminated water treated with 1% Fe<sub>2</sub>O<sub>3</sub>/SiO<sub>2</sub> nanocomposite. (i) Contaminated water treated with 1% synthetic Fe<sub>2</sub>O<sub>3</sub>/TEOS nanocomposite. (j) Contaminated water treated with 1% synthetic Fe<sub>2</sub>O<sub>3</sub>/TEOS nanocomposite.

tubules and atrophy of some glomerulus (Fig. 4 g). In addition to these histological changes, the kidney section of the rats exposed to contaminated water treated with 1%  $Fe_2O_3/SiO_2$  nanocomposite (Fig. 4 h) showed thickening of the parietal layer of Bowman's capsule and local interstitial nephritis. In a study done by Mehdi *et al.* (2021), rats exposed to 100 & 150 mg/kg silica nanoparticles showed necrosis, infiltration, dilation, and vacuolation in renal tissues and tubules with significant degrees of alterations.

The kidney section of the rats exposed to contaminated water treated with 0.5% synthetic  $Fe_2O_3$ / TEOS nanocomposite (Fig. 4 i) showed congestion of glomerular tuft and cystic dilatation of renal tubules. However, 1% synthetic Fe<sub>2</sub>O<sub>3</sub>/ TEOS nanocomposite (Fig. 4 j) showed cystic dilatation and vacuoles within tubules, without atrophy or necrosis (Boudard *et al.*, 2019).

#### CONCLUSION

Water contamination with co-existing heavy metals could be partially controlled via converting rice husk as underutilized horticultural biomass, into silica and Fe<sub>2</sub>O<sub>3</sub> silica nanocomposite using the solgel method. Silica, with its inspiring morphology and mass of 5 g, completely removed 151.73 mg Zn  $^{2+}$ , 280.20 mg Fe $^{2+}$ , and 2.35 mg Ni $^{2+}$ / L of contaminated water, as confirmed by the SEM and EDX tools. The more achievable results in this study that each 10 g mass of Fe<sub>2</sub>O<sub>3</sub> silica nanocomposite with its aggregations and Van der Waals forces showed strong affinity and selectivity to bind 40.42% of Hg<sup>2+</sup>/ L of contaminated water with coexisting heavy metals. The described solution can be more economical and environmentally alternative synthesized TEOS or Fe<sub>2</sub>O<sub>2</sub>/ TEOS than nanoparticle, which may not be sustainable and not match the health and the cost requirements. Based on a combination of serum biomarkers and histological examination for serum and renal tissues, there were no significant alterations or any indications of nephrotoxicity in rats exposed to contaminated water treated with 0.5, 1% natural silica.

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#### **Conflicts of Interest**

"The authors declare no conflict of interest."

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